

Lentiviral vectors production

Vectalys is an R&D company with a state-of-the-art technology platform for customized viral vector production.

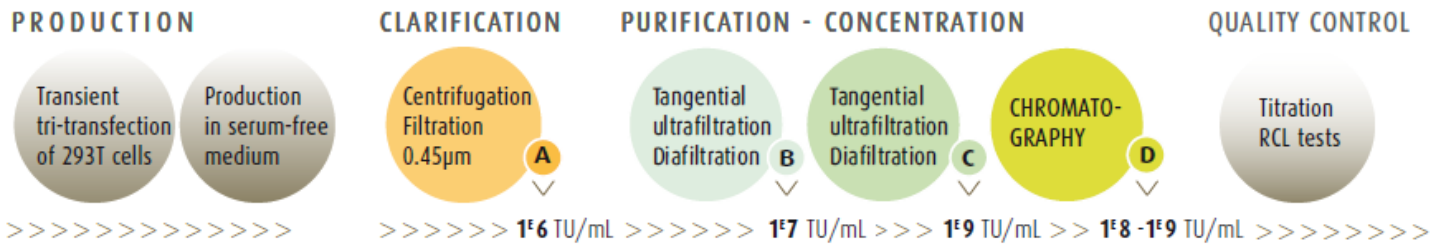
The company has developed a unique process enabling the production of high titer high purity lentiviral vectors for optimal knock-down or over expression in relevant models: primary and stem cells for target gene validation and specific tissue for animal models.

Key benefits

- 100% gene delivery achieved to primary cells such as neurons, endothelial and stem cells
- Multiple genes delivered to one cell
- High purity enabling *in vivo* injections and preventing cytotoxicity
- Dedicated support with scientific advice and biosafety assistance

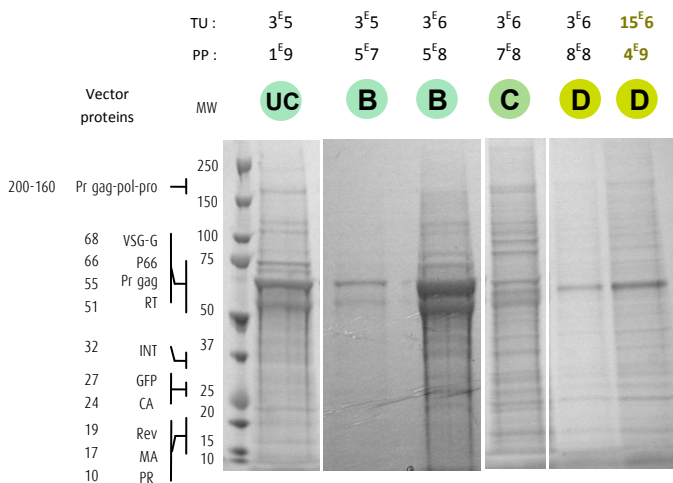
Highly concentrated and purified lentiviral vectors

Vectalys has developed a unique process for **large-scale production** of lentiviral vectors associated to concentration and purification methods for the generation of **high titer retroviral vectors**.



Our viral vector production process provides high quality lentiviral batches at different titers:

- > **High purity** : removal of major contaminants (serum and cellular proteins, plasmid and genomic DNA)
- > **High titer retroviral vectors** from **1E6 TU/mL up to 1E9 TU/mL**



High purity: Viral proteins were separated by SDS-PAGE electrophoresis and visualized by coomassie blue staining

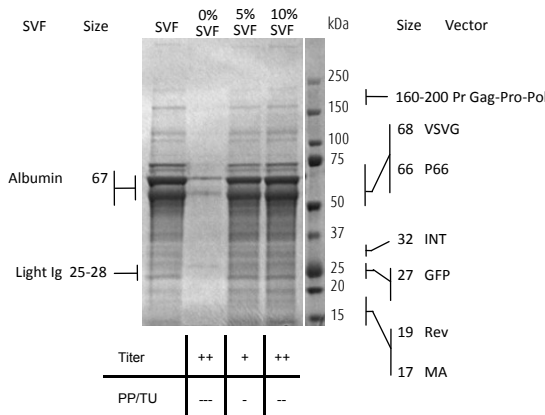
UC : classical ultracentrifugation based purification technique

	A	B	C	D
Titer (TU/mL)	2-5 ^{E6}	2 ^{E7} -3 ^{E8}	1-3 ^{E9}	1 ^{E8} -1 ^{E9}
Purity (PP/TU)	150-200	350-450	250-400	70-250
DNA yield		-10%	-70%	- 90%
Protein yield		-20%	-70%	- 95-98%
Application	Immortalized cell lines	Primary cells Stem cells		Animal models

Lentiviral vector production without serum

A robust process for the production of high titer viral supernatant in **serum free medium**:

- > improves the quality of the crude viral vectors :
 - increases the titer (TU/mL)
 - reduces the physical particles to transducing units ratio (PP/TU)
- > improves the transduction efficiency of the viral vectors



15µg of total proteins were separated by SDS-PAGE electrophoresis and visualized by coomassie blue staining

Lentiviral vector purification and concentration

Vectalys has developed **upscaleable** concentration and purification processes based on combined techniques to produce **highly concentrated** and **purified** lentiviral vectors.

Viral supernatant is purified by tangential ultrafiltration then purified and concentrated by chromatography:

- > The *ultrafiltration process* allows the concentration of viral vectors up to 2.5^{E9} TU/mL. This increases the efficiency of transduction for more delicate cells such as primary and stem cells.
- > The *chromatography* step greatly improves the purity of the final viral vector stock. These preparations are used for direct *in vivo* injection experiments.

		Concentration	Purification
Classical method	Ultracentrifugation	+++	-
	Ultrafiltration	+++	++
Vectalys process	Chromatography	-	+++
	Ultrafiltration and Chromatography	+++	++++

Applications

- > **Customized safe lentiviral vectors for your specific application**
- > **Efficient transduction of delicate cell types (i.e. primary cells, stem cells, ...)**
- > **Generation of stable cell lines**
- > **To develop *in vivo* animal models**

Quality control

- > **Production phase**
 - Standard production controls
 - Robust production cell lines
 - Drastic controls against
 - mycoplasmas
 - microbial contamination
- > **Purification and concentration phase**
 - Controlled physical parameters throughout the whole process
- > **Lentiviral batches**
 - Dedicated « viral safety rooms »
 - Standard controls of titration
 - RCR tests
 - Traceability
 - Robust technical procedures

Quality assurance is guaranteed throughout the production process

References Vergnault H. et al. **Ultra-pure retroviral production for *in vitro* and *in vivo* applications.** Human Gene Therapy, 2009, 20 (6): 671-690.

Weber A. et al. **Hepatocyte Transplantation Techniques: Large Animal Models.** In: Hepatocyte Transplantation: Methods and Protocols. Humana press, 2009, vol. 481.